

Recovery of embryos vitrified in EFS (ethylene glycol – ficoll - sucrose) solution

* Media *

Thawing solution 1 (TS1): 0.75 M sucrose in PB1 or M2 (37°C)

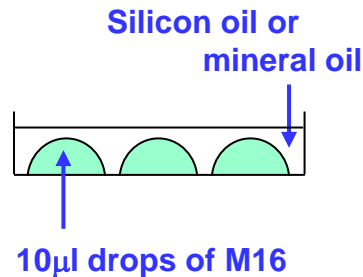
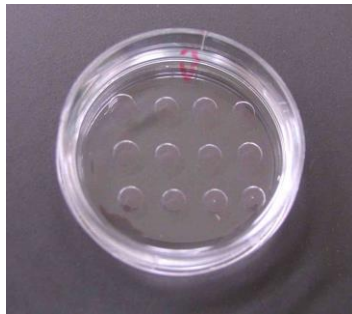
Thawing solution 2 (TS2): 0.25 M sucrose in PB1 or M2 (room temperature)

Culture medium : M16 (Sigma) containing 0.4% (w/v) BSA and equilibrated under 5% CO₂ in air at 37°C

* Equipments *

Dissecting microscope, timer, glass capillaries with mouthpieces (for handling embryos), pipettes 1,000 µl, 100 µl, watch glasses or 50 mm petri dishes (e.g., Falcon #1007), 35 mm petri dishes for embryo culture (e.g., Falcon #1008), forceps, leather cryo gloves, face mask.

Fig. 1

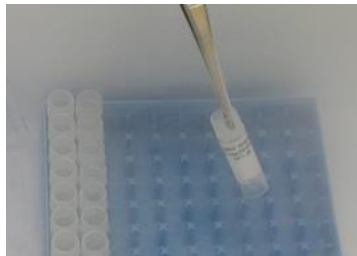


<Fig.1>

Prepare the culture dish with 10µl aliquots of M16.

Warm the TS1 to 37°C.

Fig. 2



Retrieve a tube from the liquid nitrogen.

<Fig.2>

Put on the face mask and cryogloves and retrieve a cryotube from the liquid nitrogen container using forceps.

Fig. 3



Discard the liquid nitrogen and wait for 30 s.

<Fig.3>

Quickly discard the liquid nitrogen in the tube, and let it stand for 30 sec at room temperature.

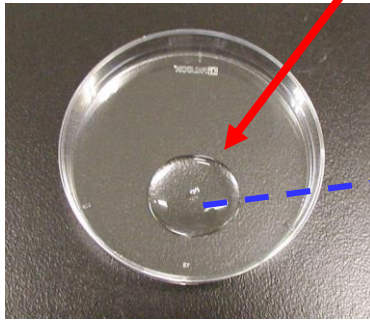
Fig. 4

Pipette 850µl of TS1 (37°C)

Mix gently by pipetting



Fig. 5



Shortly after recovery in TS1

<Fig.4>

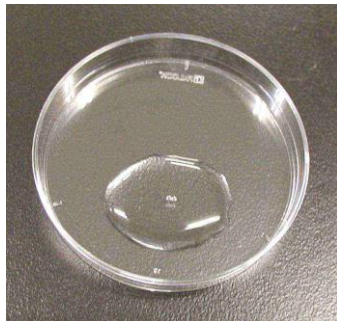
Put 850 µl of TS1 (37°C) into the tube by a single out-pipetting, and then mix the solution gently by pipetting ten times in 25 sec. Please make sure that the solution is evenly dissolved.

<Fig.5>

Using a 1000 µl pipette, transfer the entire volume to a watchglass or 50 mm petri dish. Hereafter, embryos are handled at room temperature until they are placed in M16.

Set the timer to 3 min and start.

Fig. 6



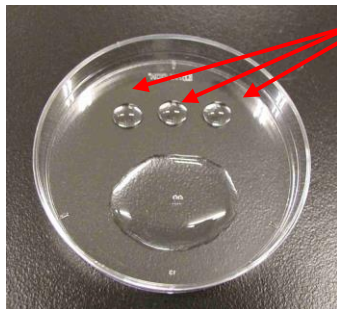
<Fig.6>

Spread the solution over the surface of the watchglass or petri dish by gentle shaking.

Dispersion of the solution makes it easier to spot the embryos.

3 min

Fig. 7

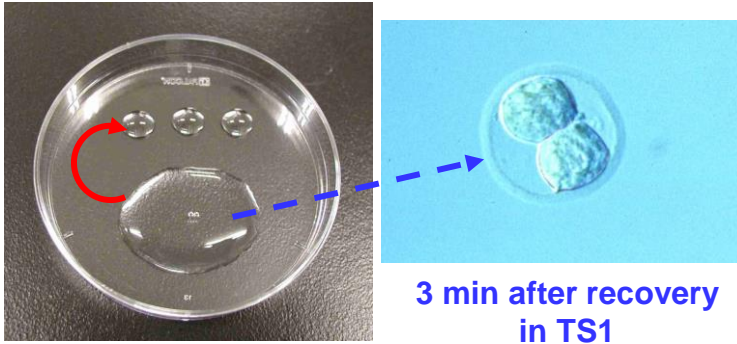


50µl drops of TS2 (room temperature)

<Fig.7>

Make three 50µl drops of TS2 onto the surface of the dish.

Fig. 8



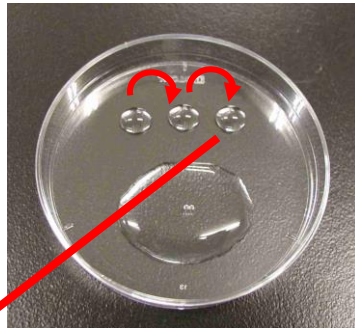
<Fig.8>

After 3 min from Fig.5, start collecting embryos using a sterile mouth-pipette, and transfer them from the drop of TS1 to a drop of TS2.

Set the timer to 3 min again and start.

3 min.

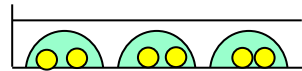
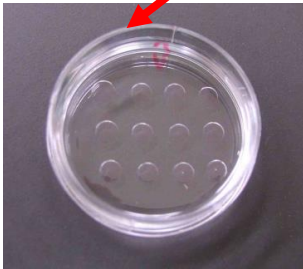
Fig. 9



<Fig.9>

Transfer the embryos to the second drop of TS2. Then, transfer them to the third drop of TS2.

Fig. 10



Culture in M16 at 37°C.

<Fig.10>

After 3 min from Fig.8, transfer the surviving embryos to 10 μ l drops of M16.

Incubate embryos under 5% CO₂ in air at 37°C until transfer to the oviducts.

Please note the followings:

- 1) The temperature of the thawing media and the duration of embryo exposure to them are critically important for the survival of embryos. Please compare the morphology of your embryos with that of embryos in the photos shown above. You may extend the period for the first thawing medium (TS1) until the embryos recover a shrunken morphology.
- 2) The embryos just after thawing are very sensitive to physical damages.
Please handle them as gently as possible.
- 3) Please thaw the first tube of embryos and make sure that the embryos finally look normal in the M16 medium. If you find any trouble with this step, please contact us.
- 4) Embryos should be transferred into day 0.5 (on the day of vaginal plug) oviducts on the day of thawing. Ideally, 16-20 embryos are to be transferred per female so that they may get pregnant efficiently.

If I can be of any further assistance, please do not hesitate to contact me.
Thank you!

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1) Preparation of BSA-free PB1

	M. W.	PB1 (BSA-free)	
		mM	mg/100ml
NaCl	58.4	136.98	800.0
KCl	74.6	2.68	20.0
KH ₂ PO ₄	136.1	1.47	20.0
Na ₂ HPO ₄ ·12H ₂ O	358.14	8.04	288.1
MgCl ₂ ·6H ₂ O	203.3	0.49	10.0
Glucose	178.6	5.56	100.0
Na pyruvate	110	0.33	3.6
CaCl ₂ ·2H ₂ O	147	0.9	13.2
Penicillin G			6.0 (approx)

Sterilize the solution by filtration.

2) Preparation of TS1 (0.75M sucrose solution)

2-1. From PB1

Dissolve 7.7 g of sucrose in PB1 (BSA free) and bring the total volume to 30 ml.

Mix by gentle shaking until sucrose is completely dissolved.

Add 90 mg of BSA onto the surface the solution and leave it stand until completely dissolved.

Sterilize the solution by filtration.

Aliquot and store in a refrigerator.

2-2. From M2

Dissolve 7.7 g of sucrose in M2 and bring the total volume to 30 ml.

Mix by gentle shaking until sucrose is completely dissolved.

Sterilize the solution by filtration.

Aliquot and store in a refrigerator.

3) Preparation of TS2 (0.25M sucrose solution)

Dilute TS1 with x2 volume of PB1 or M2, as appropriate.

Aliquot and store in a refrigerator.